

Structure Revision of Aspergillides A and B, Cytotoxic 14-Membered Macrolides from *Aspergillus ostianus*, by X-ray Crystallography

Ryuhei Ookura,¹ Keiji Kito,¹ Yota Saito,¹ Takenori Kusumi,² and Takashi Ooi*¹

¹Faculty of Pharmaceutical Sciences, The University of Tokushima, Tokushima 770-8505

²Graduate School of Science and Engineering, Tokyo Institute of Technology, Ookayama, Meguro-ku, Tokyo 152-8551

(Received January 15, 2009; CL-090059; E-mail: tooi@ph.tokushima-u.ac.jp)

m-Bromobenzoates of aspergillides A and B gave crystals suitable for X-ray crystallography, resulting in the revision of the structures of both compounds as **1** and **3**, respectively.

We have reported the structures of aspergillides A, B, and C, which were obtained from marine-derived fungus *Aspergillus ostianus* and which each had a unique 14-membered macrolactone ring.¹ The structures of aspergillides A and B were proposed to be **3** and **5**, respectively, based on NMR analyses including NOEs and ¹H, ¹H-coupling constants.¹ Recently, Hande and Uenishi succeeded in the total synthesis of a compound possessing the proposed structure **3**, and found that its NMR data were different from that of aspergillide A but identical to that of aspergillide B.² Therefore, the structure of aspergillide B ought to be revised to **3**. This change throws the real structure of aspergillide A into question.

In our previous study, the absolute configurations at the 4- and 13-positions of aspergillide A had been firmly established by the modified Mosher's method³ carried out for the compound itself (C-4) and its methanolysis product (C-13). Thus, our most likely error was incorrect assignment of the stereochemistry at C-3 and C-7.

In order to reexamine the structures of aspergillides A and B, we performed a large-scale (45 L) cultivation of the fungus again, and obtained two compounds, labeled I (9.2 mg) and II (48.5 mg), whose physical properties (NMR, TLC, and HPLC) were identical to those of aspergillides A and B, respectively.

The *p*-bromobenzoate of II failed to crystallize, although the *m*-bromobenzoates⁴ of I and II afforded nice crystals suitable for X-ray crystallography. The results are shown in Figure 1. The X-ray structures, **2**-X-ray (I-*m*-bromobenzoate) and **4**-X-ray (II-*m*-bromobenzoate), unambiguously established the absolute stereochemistry of aspergillides A (**1**) and B (**3**). The latter structure nat-

urally accords with the one revised by Hande and Uenishi,² and the former one has an unexpected *cis* bridge at C-3 and C-7.

Since the correct structures **1** and **3** were obtained, we rechecked the previous experiments for aspergillides A and B to clarify the causes of the mistakes. First, we had assigned a small peak appearing at the cross section of H-3 and H-8 as a real cross peaks between the protons and overlooked the cross peaks between H-2a and H-8, which were relevant to the *cis* juncture at C-3 and C-7 in the NOESY spectrum (400 MHz, CDCl₃)¹ of aspergillide A, leading to the *trans* bridge structure **3**. However, since the chemical shift of H-3 (δ 4.23) was very close to that of H-7 (δ 4.21), it could be a cross peak between H-8 and H-7. Change of the solvent to DMSO-*d*₆ resulted in satisfactory separation of the signals [H-7 (δ 4.03) and H-3 (δ 3.96)], and distinct NOE cross peaks were observed between the protons.⁵ The cross peaks between H-8 (δ 5.88) and H-2a (δ 2.55) were also observed. Secondly, although the absolute stereochemistry at C-4 of aspergillide B (previously proposed as **5**) had been determined by the modified Mosher's method, that at C-13 was assumed, without an experiment, to be opposite to that at C-13 of aspergillide A (previously proposed as **3**), because the stereochemistry at C-3, -4, and -7 of both compounds had been thought to be the same.

With respect to structure **6** of aspergillide C, the absolute configurations as C-4 and -13 had been established by the modified Mosher's method.¹ Moreover, because of the presence of C₅=C₆ in **6**, separation of the ¹H NMR signals was much better than that observed for aspergillides A and B, we were able to reconfirm the *trans* stereochemistry at C-3 and C-7 of **6**.

Our comparison of the structures of aspergillides A (**1**) and B (**3**) led us to wonder whether or not the two compounds could be equilibrated via a *retro*-Michael reaction (cleavage of C₃–O forming C₂=C₃). Allowing **3** on silica gel to stand for 24 h did not yield a trace of **1**. Reaction of **1** with sodium methoxide afforded a methanolysis product as a single isomer.¹ Therefore, it seems most likely that both compounds are genuine natural products.

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References and Notes

- 1 K. Kito, R. Ookura, S. Yoshida, M. Namikoshi, T. Ooi, T. Kusumi, *Org. Lett.* **2008**, *10*, 225.
- 2 S. M. Hande, J. Uenishi, *Tetrahedron Lett.* **2009**, *50*, 189.
- 3 I. Ohtani, T. Kusumi, Y. Kashman, H. Kakisawa, *J. Am. Chem. Soc.* **1991**, *113*, 4092.
- 4 Crystallographic data reported in this manuscript have been deposited with Cambridge Crystallographic Data Centre as supplementary publication numbers CCDC-710343 for **2** and -710344 for **4**. Copies of the data can be obtained free of charge via www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge, CB2 1EZ, UK; fax: +44 1223 336033; or deposit@ccdc.cam.ac.uk).
- 5 Supporting Information is also available electronically on the CSJ-Journal Web site, <http://www.csj.jp/journals/chem-lett/index.html>.

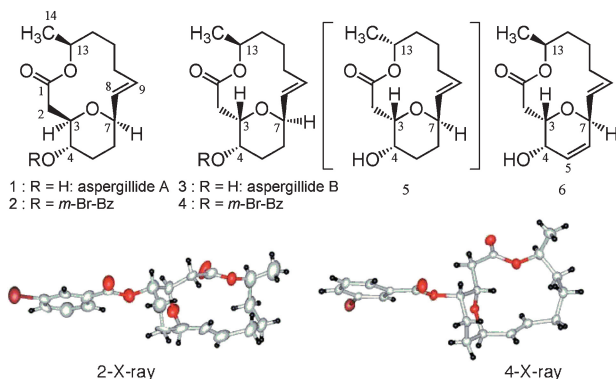


Figure 1. (above) Correct structures of aspergillides A (**1**) and B (**3**) and incorrect structure **5** reported as aspergillide B. Structure **6** of aspergillide C was reconfirmed in this study. (below) X-ray structures of **2** and **4**.